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Adsorption of Sodium Dodecyl Sulfate onto Activated Coconut Shell-Based Adsorbent: Isothermal Remodelling

Shakirah Abdul Wahab Sha'arani¹, Nur Adeela Yasid¹, Mohd Izuan Effendi Halmi², Ahmad Razi Othman³ and Mohd Yunus Shukor¹*

¹Department of Biochemistry, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia, 43400 UPM Serdang, Selangor, Malaysia.

²Department of Soil Management, Faculty of Agriculture Universiti Putra Malaysia, 43400 UPM Serdang, Selangor, Malaysia.

³Department of Chemical Engineering, Faculty of Engineering and Built Environment, Universiti Kebangsaan Malaysia, 43600 UKM Bangi, Selangor, Malaysia.

> *Corresponding author: Mohd Yunus Shukor, Department of Biochemistry, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia, 43400 UPM Serdang, Selangor, Malaysia.

Email: mohdyunus@upm.edu.my

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ABSTRACT

A remodelling evaluation was conducted on the sorption isotherm data for SDS adsorption onto activated coconut shell using nonlinear regression. A total of nineteen models, including BET, Brouers-Sotolongo, Dubinin-Radushkevich, Fowler-Guggenheim, Freundlich, Fritz-Schlunder III, Hill, Henry, Jovanovic, Khan, Langmuir, Moreau, Radke-Prausnitz, Redlich-Peterson, Sips, Temkin, Toth, Unilan, and Vieth-Sladek, were employed to determine the best fit through nonlinear regression. All models were found to exhibit good fits to the data, except for the Fowler-Guggenheim, Henry and Dubinin-Radushkevich models. Statistical analysis based on error function assessments, including accuracy factor (AF), bias factor (BF), root-mean-square error (RMSE), adjusted coefficient of determination (adjR²), Bayesian Information Criterion (BIC), corrected AICc (Akaike Information Criterion), and Hannan-Quinn Criterion (HQC), revealed that the best performance was achieved by the Moreau model followed by (descending order) Unilan, Redlich-Peterson, Fritz-Schlunder III, Toth, and Langmuir. The maximum adsorption capacity estimates given by the Moreau and Langmuir models were better in line with experimental findings. The value of the maximum monolayer adsorption capacity for SDS binding to activated coconut shell according to the Langmuir's parameter q_{mL} was 81.93 mg g⁻¹ (95% Confidence interval from 76.422 to 87.440), while b_L (L mg⁻¹), the Langmuir model constants was 0.10 L mg⁻¹ (95% C.I. from 0.070 to 0.133). The value of the maximum monolayer adsorption capacity for SDS binding to activated coconut shell according to the Moreau's parameter q_{mM} was 90.82 mg g⁻¹ (95% Confidence interval from 76.336 to 105.303), while b (L mg⁻¹), the Moreau's model constant was 0.15 L mg⁻¹ (95% C.I. from 0.089 to 0.203) and l, another Moreau's dimensionless constant was 0.3 (95% C.I. from -0.049 to 0.646).

INTRODUCTION

Surfactants are widely utilized in both household and industrial settings owing to their typical physicochemical properties, particularly their surface activity. These molecules consist of a hydrophilic head and a hydrophobic tail, which give them amphiphilic properties [1]. Surfactants are categorized into four types based on their chemical structure: cationic, anionic, nonionic, and zwitter ionic [2]. Anionic surfactants serve as pivotal components in various products such as detergents, toothpaste, shampoo, pesticides, textile paints and personal care items [3,4]. Moreover, they find application in diverse fields including soil remediation, enhanced oil recovery, dispersion, emulsion polymers, deinking, ore flotation and cutting oils [5]. Because of their extensive use and versatility, surfactants frequently end up in both industrial and domestic wastewater, as they might not be fully utilized in their intended applications. Some of the discharged surfactants may undergo natural degradation. However, reliance solely on biodegradation or natural breakdown might not suffice, especially when surfactants are present in high concentrations [6]. Residual surfactants, particularly the anionic variants, can adversely affect aquatic microorganisms and ecosystems in water bodies. Additionally, if these surfactants enter the food chain, they can also pose risks to humans [7]. Moreover, the presence of surfactants can impact the effectiveness of wastewater treatment plants [8]. For instance, surfactants can influence sludge dewatering characteristics and enhance the solubility of organic compounds, thus affecting treatment efficacy [9]. Therefore, it is crucial to devise an efficient treatment method for removing surfactants, as attempted in the current study, focusing on a specific case of SDS.

SDS, a fundamental and significant anionic surfactant, finds extensive application in detergents, shampoos, and cosmetics. SDS has been linked to symptoms like depression, difficulty breathing, and diarrhea in both humans and animals [10]. SDS also induces acute and chronic toxicity in fish [11]. Surfactants elevate the solubility of organic compounds in water, potentially increasing the carcinogenicity and risk of dermatitis, as documented in literature [7,12]. Given the environmental and health concerns, the acceptable limits for surfactants in water intended for domestic use are set at 1 mg/L, and even lower at 0.5 mg/L for potable water [13]. These stringent regulations underscore the necessity for treating wastewater containing surfactants. Thus, the significance of the present study on SDS removal is highlighted.

Several methods, including adsorption [2,4,5,13], ultra-filtration [14,15], biodegradation [10] and advanced oxidation [9], are commonly used for wastewater treatment. Among these methods, adsorptive separation stands out due to its versatility in fine-tuning adsorbent performance, the potential for synthesizing adsorbents from biomass or waste materials, and its feasibility for large-scale continuous operation. The synthesis of adsorbents from various materials, including biomass waste such as coconut shell [5,16], wood, wood stalks, sawdust, sugarcane bagasse, seeds, seed hulls, jackfruit peels, coffee beans [17-21], microalgae, etc., has been extensively documented in the literature. In previous study [16], the adsorptive separation of sodium dodecyl benzene sulfonate (SDBS) using a biosorbent derived from coconut shells was investigated, demonstrating its efficacy. The current research extends the work by applying the same biosorbent (PAACS-I) to the adsorptive separation of SDS [5], another significant anionic surfactant. The influence of various operating parameters on the SDS removal efficiency was examined, conducting a comprehensive analysis of kinetics and thermodynamics to elucidate the adsorption mechanism and heat effects.

Gaining a full understanding of the biosorption process in distinct species requires a careful examination of the kinetics and isotherms. The data in question is intrinsically nonlinear, although scientific publications frequently depict it as though it were linear. Nevertheless, the error structure linked to essentially nonlinear data is changed during the linearization process. One problem with this method is that it makes the modified data's residuals seem less likely to follow a normal distribution. [22]. Consequently, it gets increasingly difficult to put a number on uncertainty, which is typically shown as a 95% confidence range. The purpose of this research is to review and reevaluate an earlier paper on SDS sorption upon AC shroud [5], which employed linear regression to derive the best-fitting models.

METHOD

Data acquisition and fitting

Figure 9 data (323 K) from a previously published study [5] was digitized using the freeware Webplotdigitizer 2.5 [23]. The program's digitization capabilities have garnered accolades for their

reliability [24]. Following that, Curve-Expert Professional (Version 2.6.5, copyright Daniel Hyams), a program for curve fitting, was used to perform nonlinear regression on the data. The software package MATLAB (Mathworks, Massachusetts, USA) was used to resolve the implicit equations.

Isotherms

As the value of the data points is very small, only models having parameters of up to three were considered to prevent overfitting.

Table 1. Mathematical models that were used in modelling data.

| Isotherm | n | Formula | Pof | | | | | |
|---|---------------|--|---------|--|--|--|--|--|
| Henry's law | <i>p</i> 1 | $q_{e} = HC_{e}$ | [25] | | | | | |
| Langmuir | 2 | $q_e = \frac{q_{mL}b_L C_e}{q_{mL}b_L C_e}$ | [26] | | | | | |
| Jovanovic | 2 | $q_e = q_{ml} (1 - e^{-K_j C_e})$ | [27] | | | | | |
| Freundlich | 2 | $a = K_{-} C^{\frac{1}{n_{F}}}$ | [28] | | | | | |
| Dubinin-Radush- | 2 | $q_e = n_F \sigma_e$ | [29,30] | | | | | |
| kevich | | $a = a \qquad \exp\left\{-\kappa \left[pT\ln\left(1+\frac{1}{2}\right)\right]^2\right\}$ | | | | | | |
| | | $q_e = q_{mDR} exp\left\{-\kappa_{DR}\left[\kappa_{III}\left(1 + \frac{1}{C_e}\right)\right]\right\}$ | [21 22] | | | | | |
| | | correct form $\left(\left[\left[\left(\left(c \right) \right)^{2} \right]^{2} \right) \right)$ | [31,32] | | | | | |
| | | $q_e = q_{mDR} exp \left\{ -K_{DR} \left[RT ln \left(\left(\frac{C_s}{C_e} \right) \right) \right] \right\}$ | | | | | | |
| Tambin | 2 | $a = \frac{RT}{(lm(a, f))}$ | [33,34] | | | | | |
| Temkin | 3 | $q_e = \frac{1}{b_T} \{ ln(a_T c_e) \}$ | [25] | | | | | |
| Redlich-Peterson | 3 | $q_e = \frac{K_{RP1} C_e}{1 + K_{RP2} C_e^{\beta_{RP}}}$ | [33] | | | | | |
| | | $K = \frac{1}{n_s}$ | [36] | | | | | |
| Sips | 3 | $q_e = \frac{n_s q_{ms} c_e}{\frac{1}{n_s}}$ | | | | | | |
| | | $1 + K_s C_e^{ns}$ $a_m \tau C_e$ | [37] | | | | | |
| Toth | 3 | $q_e = \frac{m_1}{\left(K_T + C_e^{n_T}\right)^{n_T}}$ | [*,] | | | | | |
| Hill | 3 | $q_e = \frac{q_{mH} C_e^{n_H}}{q_{e} - q_{e} - q_$ | [38] | | | | | |
| | | $K_H + C_e^{H}$ $q_{mK}b_K C_e$ | [39] | | | | | |
| Khan | 3 | $q_e = \frac{1}{(1 + b_K C_e)^{a_K}}$ | [40] | | | | | |
| BET | 3 | $q_e = \frac{q_{mBET} \alpha_{BET} c_e}{(1 - \beta_{BET} c_e)(1 - \beta_{BET} c_e + \alpha_{BET} c_e)}$ | [40] | | | | | |
| Vieth-Sladek | 3 | $q_e = \frac{q_{mVS}b_{VS}C_e}{(1+b-C)^{n_{VS}}}$ | [41] | | | | | |
| Dadlas Dromanita | 2 | $A_{RP}B_{RP}C_e^{\beta}$ | [42-44] | | | | | |
| Rauke-Frausintz | 3 | $q_e = \frac{1}{A_{RP} + B_{RP}C_e^{\beta - 1}}$ | | | | | | |
| Brouers-Sotolongo | 3 | $q_e = q_{mBS} \left[1 - \left(1 + (0.5) \left(\frac{t}{\tau} \right)^{\alpha} \right)^{-2} \right]$ | [45,46] | | | | | |
| Fritz-Schlunder-III | 3 | $a = \frac{q_{mFS}K_{FS}C_e}{q_{mFS}K_{FS}C_e}$ | [47] | | | | | |
| Fowler-Guggen- | 5 | $q_e = 1 + K_{FS}C_e^{n_{FS}}$ | [48] | | | | | |
| heim* | 3 | $K_{I}C_{a}e^{\frac{\alpha q_{e}}{q_{mFG}}}$ | [10] | | | | | |
| | 5 | $q_e = q_{mFG} \frac{\alpha q_e}{1 + K_F C_e e^{\frac{\alpha q_e}{q_{mFG}}}}$ | | | | | | |
| Moreau | 3 | $bC_e + lb^2 C_e^2$ | [49] | | | | | |
| | 5 | $q_e - q_{mM} \frac{1}{1 + 2bC_e + lb^2C_e^2}$ | | | | | | |
| Unilan | 3 | $q_e = \frac{q_{mU}}{2b_{II}} \ln\left(\frac{a_U + c_e e^{-b_U}}{a_{II} + c_e e^{-b_U}}\right)$ | | | | | | |
| Baudu | 4 | $a = \frac{q_{mB}b_BC_e^{(1+x+y)}}{q_{mB}b_BC_e^{(1+x+y)}}$ | [50] | | | | | |
| Buudu | | $q_e = 1 + b_B C_e^{(1+x)}$ | [51] | | | | | |
| Marczewski-Jaroniec | : 4 | $a_{i} = a_{mMI} \left(\frac{\left(K_{MJ} C_{e} \right)^{n_{MJ}}}{\left(K_{MJ} C_{e} \right)^{n_{MJ}}} \right)^{\frac{m_{MJ}}{n_{MJ}}}$ | [21] | | | | | |
| | | $q_e - q_{mMJ} \left(1 + \left(K_{MJ} C_e \right)^{n_{MJ}} \right)$ | [47] | | | | | |
| Fritz-Schlunder-IV | 4 | $q_e = \frac{A_{FS}C_e^{PFS}}{1 + B_{e-s}C_{FS}^{b_{FS}}}$ | [4/] | | | | | |
| Weber-van Vliet* | 4 | $C = P_{e,q} \left(\frac{P_{2}q_{e}^{P_{3}} + P_{4}}{P_{2}q_{e}^{P_{3}} + P_{4}} \right)$ | [52] | | | | | |
| $c_e = F_1 q_e$ Note *Implicit equation or function. | | | | | | | | |

Statistical analysis

The following statistical discriminatory tests were utilized in this study: HQ, BF, AF, RMSE, adjusted coefficient of determination (R^2), corrected Akaike Information Criterion (AICc), BIC, and Hannan and Quinn's Criterion (AIC). The root-mean-squared error (RMSE) was calculated using Eqn. 1, which indicates that in most cases, a reduced RMSE is obtained with fewer parameters. Where n is the total number of observations, Obi and Pdi are the predicted and observed values, and p is the total number of parameters utilized in this context [22].

$$RMSE = \sqrt{\frac{\sum_{i=1}^{n} (Pd_i - Ob_i)^2}{n-p}}$$
(Eqn. 1)

The modified R^2 is utilized to overcome the limitation that R^2 , which stands for the coefficient of determination, does not take into account the total number of parameters in a model. The total variance of the y-variable is represented by S^2_y in the equation (Equations 2 and 3), while RMS refers to the Residual Mean Square.

Bias factor =
$$10\left(\sum_{i=1}^{n} \log \frac{(Pd_i/Ob_i)}{n}\right)$$
 (Eqn. 2)

Accuracy factor = $10\left(\sum_{i=1}^{n} \log \frac{|(Pd_i/Ob_i)|}{n}\right)$ (Eqn. 3)

In the AICc equation, p is the number of parameters and n is the number of data points, and the AICc is derived accordingly. Data sets with many parameters but few values are best handled by applying the updated Akaike information criterion (AICc). [53]. It is more probable that a model is correct if its AICc score is lower [53]. The AIC's (Akaike Information Criterion) basis is information theory. It finds a happy medium between a model's complexity and its goodness of fit [54].

$$AICc = 2p + n \ln\left(\frac{RSS}{n}\right) + \frac{2(p+1) + 2(p+2)}{n - p - 2}$$
(Eqn. 4)

In addition to AICc, the Bayesian Information Criterion (BIC) (Equation 5) is another information-theory-based statistical tool. This error function penalizes parameter count more harshly than AIC [55].

$$BIC = n \ln\left(\frac{RSS}{n}\right) + k \ln(n)$$
 (Eqn. 5)

Equation 6 represents the Hannan-Quinn information criterion (HQC), an additional information theory-based method for error functions. The HQC is more reliable than the AIC due to the inclusion of the $\ln n$ element in the calculation [53].

$$HQC = nIn\left(\frac{RSS}{n}\right) + 2kIn(In n)$$
(Eqn. 6)

Two further error function analyses extracted from Ross's research are the Accuracy Factor (AF) and the Bias Factor (BF) [53]. Rather than penalizing models based on their parameter counts, these error functions empirically assess their goodnessof-fit (**Equations 7** and **8**).

Adjusted (
$$R^2$$
) = $1 - \frac{RMS}{S_Y^2}$ (Eqn. 7)

Adjusted
$$(R^2) = 1 - \frac{(1-R^2)(n-1)}{(n-p-1)}$$
 (Eqn. 8)

A typical error function utilized in much isotherm research is Marquardt's percent standard deviation (MPSD). Depending on the number of degrees of freedom in the system, the function takes on a shape similar to a geometric mean error distribution [56]. Among the first to use this error function in the adsorption field is [57] and the McKay group first proposed the official name for the error function, which is MPSD (Equation 9) [58].

$$MPSD = 100 \sqrt{\frac{1}{n-p} \sum_{i=1}^{n} \left(\frac{Ob_i - Pd_i}{Ob_i}\right)^2}$$
(Eqn. 9)

in which n is the total number of experimental data points, p is the total number of parameters, *Obi* is the set of experimental data points, and *Pdi* is the value that the model predicts.

RESULTS AND DISCUSSION

Several models were applied to the equilibrium data given in [5] by non-linear regression. Importantly, as shown in **Figs.** 1–18, all of these models showed good data fits with the exception of the Henry, Dubinin–Radushkevich models. There was no convergence in the Fowler-Guggenheim model. Among several assessment metrics, the Moreau isotherm model stood out as the top performer. These metrics included having the shortest RMSE, adjusted R², AF, and BF values close to one, as well as the lowest values for Bayesian Factor (BF), MPSD, and AICc values. Nevertheless, the Langmuir model emerged as the top performer when AICc was utilized as the only error function. Moreau, Unilan, Redlich-Peterson, Fritz-Schlunder III, Toth, and Langmuir were the top models in descending order according to the majority error function analysis.

The Unilan, Redlich-Peterson, Fritz-Schlunder III, and Toth models all performed well, but their maximum adsorption capacities were very different from the experimental values, and the wide 95% confidence interval demonstrated that they were not a good fit. The small sample size is probably to blame for this disparity. On the other hand, the maximum adsorption capacity estimates given by the Moreau and Langmuir models were better in line with experimental findings. Nonlinear regression provides more accurate results than linear regression, which was utilized in the original publication—which only presented the linearized forms of the Langmuir, Temkin, Dubinin-Radushkevich, and Freundlich models—because many models fit the bentonite data well.



Fig. 1. SDS adsorption isotherm onto activated coconut shell as modelled using the Henry model.



Fig. 2. SDS adsorption isotherm onto activated coconut shell as modelled using the Langmuir isotherm model.



Fig. 3. SDS adsorption isotherm onto activated coconut shell as modelled using the Freundlich isotherm model.



Fig. 4. SDS adsorption isotherm onto activated coconut shell as modelled using the Temkin isotherm model.



Fig. 5. SDS adsorption isotherm onto activated coconut shell as modelled using the Dubinin-Radushkevich isotherm model.



Fig. 6. SDS adsorption isotherm onto activated coconut shell as modelled using the Jovanovic isotherm model.



Fig. 7. SDS adsorption isotherm onto activated coconut shell as modelled using the Redlich-Peterson isotherm model.



Fig. 8. SDS adsorption isotherm onto activated coconut shell as modelled using the Sips isotherm model.



Fig. 9. SDS adsorption isotherm onto activated coconut shell as modelled using the Toth isotherm model.



Fig. 10. SDS adsorption isotherm onto activated coconut shell as modelled using the Hill isotherm model.



Fig. 11. SDS adsorption isotherm onto activated coconut shell as modelled using the Khan isotherm model.



Fig. 12. SDS adsorption isotherm onto activated coconut shell as modelled using the BET isotherm model.



Fig. 13. SDS adsorption isotherm onto activated coconut shell as modelled using the Vieth-Sladek isotherm model.



Fig. 14. SDS adsorption isotherm onto activated coconut shell as modelled using the Radke-Prausnitz isotherm model.



Fig. 15. SDS adsorption isotherm onto activated coconut shell as modelled using the Brouers-Sotolongo isotherm model.



Fig. 16. SDS adsorption isotherm onto activated coconut shell as modelled using the Fritz-Schlunder III isotherm model.



Fig. 17. SDS adsorption isotherm onto activated coconut shell as modelled using the Unilan isotherm model.



Fig. 18. SDS adsorption isotherm onto activated coconut shell as modelled using the Moreau isotherm model.

Table 2. Error function analysis for the fitting of the isotherm of SDS onto activated coconut shell.

| Model | MPSD | RMSE | adR2 | AICc | BIC | HQC | BF | AF |
|-----------------------------|--------|-------|------|-------|-------|-------|------|------|
| Moreau | 157.5 | 1.58 | 1.00 | 36.45 | 8.28 | 6.44 | 1.00 | 1.01 |
| Unilan | 158.8 | 1.59 | 1.00 | 36.56 | 8.40 | 6.55 | 1.00 | 1.01 |
| Redlich-Peterson | 159.7 | 1.60 | 1.00 | 36.63 | 8.47 | 6.63 | 1.00 | 1.01 |
| Fritz-Schlunder III | 159.7 | 1.60 | 1.00 | 36.63 | 8.47 | 6.63 | 1.00 | 1.01 |
| Toth | 159.9 | 1.60 | 1.00 | 36.66 | 8.49 | 6.65 | 1.00 | 1.01 |
| Khan | 160.3 | 1.60 | 1.00 | 36.69 | 8.52 | 6.68 | 1.00 | 1.01 |
| Radke-Prausnitz | 160.3 | 1.60 | 1.00 | 36.69 | 8.52 | 6.68 | 1.00 | 1.01 |
| Hill | 160.7 | 1.61 | 1.00 | 36.73 | 8.56 | 6.72 | 1.00 | 1.02 |
| Sips | 160.7 | 1.61 | 1.00 | 36.73 | 8.56 | 6.72 | 1.00 | 1.02 |
| Brouers-Sotolongo | 162.0 | 1.62 | 1.00 | 36.84 | 8.68 | 6.84 | 1.00 | 1.02 |
| Vieth-Sladek | 174.2 | 1.74 | 0.99 | 37.85 | 9.69 | 7.85 | 1.00 | 1.01 |
| BET | 177.9 | 1.78 | 0.99 | 38.15 | 9.98 | 8.14 | 1.00 | 1.01 |
| Marczewski | 183.3 | 1.83 | 0.99 | 80.55 | 10.34 | 7.88 | 1.00 | 1.01 |
| Baudu | 184.2 | 1.84 | 0.99 | 80.62 | 10.40 | 7.94 | 1.00 | 1.01 |
| Fritz-Schlunder IV | 184.2 | 1.84 | 0.99 | 80.62 | 10.40 | 7.94 | 1.00 | 1.01 |
| Weber-van Vliet | 187.6 | 1.88 | 0.99 | 80.88 | 10.66 | 8.21 | 1.00 | 1.02 |
| Temkin | 190.9 | 1.91 | 0.99 | 39.13 | 10.97 | 9.13 | 1.00 | 1.02 |
| Langmuir | 240.9 | 2.41 | 0.99 | 27.95 | 13.84 | 12.61 | 1.00 | 1.03 |
| Freundlich | 312.4 | 3.12 | 0.98 | 31.59 | 17.48 | 16.25 | 1.01 | 1.04 |
| Jovanovic | 525.2 | 5.25 | 0.96 | 38.87 | 24.76 | 23.53 | 0.98 | 1.07 |
| Dubinin-Radush- | | | | | | | | |
| kevich | 2577.8 | 25.78 | 0.39 | 61.14 | 47.03 | 45.80 | 0.22 | 4.86 |
| Henry | 2643.2 | 26.43 | 0.43 | 53.77 | 46.71 | 46.10 | 0.57 | 1.92 |
| Fowler-Guggen- | | | | | | | | |
| heim | n.a. | n.a. | n.a. | n.a. | n.a. | n.a. | n.a. | n.a. |
| Note: | | | | | | | | |
| RMSE Root mean Square Error | | | | | | | | |

Adjusted Coefficient of determination adR

no of parameters

ÅF Accuracy factor BF

Bias factor

BIC Bayesian Information Criterion AICc Adjusted Akaike Information Criterion HQC Hannan–Quinn information criterion

Moreau

Interactions between adsorbates occur on a homogenous adsorbent surface, as described by the Moreau isotherm model [59], furthermore, the Ruthven model, which was created for zeolite adsorption, is very similar to this one [60]. Proteins adsorb to polymer surfaces by a complex web of interactions, some of which include hydrogen bonding, van der Waals forces, and hydrophobic effects. Taking into account the heterogeneity and interaction among adsorbed proteins, the Moreau isotherm was determined to be the most effective model for the adsorption process [61]. The value of the maximum monolayer adsorption capacity for SDS binding to activated coconut shell according to the Moreau's parameter q_{mM} was 90.82 mg g⁻¹ (95% Confidence interval from 76.336 to 105.303), while b (L mg⁻¹), the Moreau's model constant was 0.15 L mg⁻¹ (95% C.I. from 0.089 to 0.203) and l, another Moreau's dimensionless constant was 0.3 (95% C.I. from -0.049 to 0.646).

Unilan (Unilin) model

Among the empirical correlations proposed for use in equilibrium data analysis in the book by Valenzuela and Myers is Unilan. Some publications also use the name Unilin when UniLan is more appropriate. An acronym for "Uniform distribution and Langmuir local" isotherm, UniLan describes this design. Assuming a surface that is defined patch-wise, the Unilan equation states that the local Langmuir equation is applicable to each patch [62]. The Unilan equilibrium constant (a_U) and model exponent (*bU*) are used in this isotherm, while $q_{mU} (\text{mg g}^{-1})$ is the maximal monolayer adsorption capacity predicted by the isotherm.

The variability of the system is described by b_U . The more this parameter's value increases, the more diverse the system gets. The UniLan equation is transformed into the classical Langmuir equation when $b_U = 0$ in this limit, which also happens when the value for the range of energy distribution becomes zero [63]. The result of the remodeling exercise, however, did not conform to the observed experimental q_m value of about 90 mg g⁻¹ (**Table 3**).

Langmuir isotherm

There is a continuum between empirical and mechanistic isotherm models; the Langmuir isotherm is definitely in the former. On the assumption of structural homogeneity in the adsorbent and equal energy for all adsorption sites, this model postulates that adsorption occurs as a uniform monolayer on the adsorbent surface [64]. Because intermolecular interactions decrease exponentially with increasing distance, it predicts that a monolaver will form on the outer adsorbent surface. The linearization of the relationship and the assumption of a constant monolayer adsorption capacity are both brought about by this. That Henry's model holds true for both extremely dilute and concentrated solute concentrations is another important finding [65].

The Freundlich and Langmuir models are two of the most popular choices for sorption research. The nonlinear regression method was able to get parameter estimates that were quite similar to the original study's, but it couldn't give a 95% confidence interval. The value of the maximum monolayer adsorption capacity for SDS binding to activated coconut shell according to the Langmuir's parameter qmL was 81.93 mg g⁻¹ (95% Confidence interval from 76.422 to 87.440), while b_L (L mg⁻¹), the Langmuir model constants was 0.10 L mg⁻¹ (95% C.I. from 0.070 to 0.133). These values are very similar to the reported linear regressed values in the original publication for q_{mL} and b_L values at 90.9 mg g⁻ 1 and 0.09 L mg⁻¹, respectively [5] (Table 4).

Toth isotherm

The Toth isotherm model serves as an equation used to explain adsorption processes, on surfaces with variations. It is crafted to address differences from the Langmuir isotherm, which assumes adsorption sites and single layer adsorption. The Toth isotherm provides flexibility in describing adhesion behaviors that differ from the Langmuir model by accommodating both high concentrations of adsorbate. By incorporating the parameter n_T it accommodates surface diversity simplifying to match the Langmuir isotherm. In fields such, as engineering, wastewater treatment and material science the process of adsorption involves capturing pollutants, dyes, heavy metals and other impurities on different materials like activated carbon, clays and biochar.

The Toth isotherm is preferred over models as it fits data better especially in cases of varied adsorption environments. This model can handle a range of adsorption scenarios due to its parameters. However it is more intricate than models like Langmuir or Freundlich as it requires estimating parameters. The Toth isotherm serves as an adaptable tool for explaining adsorption on surfaces with greater precision for systems that deviate from simpler model assumptions. The Toth isotherm was found as the best isotherm for the adsorption of two organic dyes (alizarin red S and phenol red) on mesoporous silica and hybrid gels [66], the adsorption of the red, yellow, and mustard yellow acid dyes onto granular activated carbon [67] adsorption of CO₂ using biochar and KOH and ZnCl2 activated carbons derived from pine sawdust [68] and adsorption of ammonia onto an activated carbon [69].

Redlich-Peterson isotherm

The Redlich-Peterson isotherm is a widely used model in adsorption studies that combines elements of both the Langmuir and Freundlich isotherms. It is particularly valuable for describing adsorption processes on heterogeneous surfaces where the adsorption energy varies. The model's flexibility stems from the exponent β_{RP} , which allows it to transition between the Langmuir isotherm (when $\beta_{RP}=1$) and the Freundlich isotherm (when β_{RP} approaches 0). This makes it suitable for describing adsorption on both homogeneous and heterogeneous surfaces. Unlike simpler isotherm models, the Redlich-Peterson isotherm accounts for the non-linearity of adsorption processes. This can provide a more accurate fit for experimental data, especially when the adsorption sites have varying affinities. The model is extensively used in environmental engineering, chemical engineering, and materials science for the study of adsorption phenomena. It is applied in the removal of contaminants from water, adsorption of gases on solids, and in the design of adsorption systems for industrial processes.

The parameters are usually estimated using non-linear regression techniques. Accurate estimation of these parameters is crucial for predicting the adsorption capacity and understanding the adsorption mechanism. While the Redlich-Peterson isotherm provides a good fit for a wide range of data, its complexity can be a drawback. The need for non-linear regression and the interpretation of three parameters can complicate its application compared to simpler models. In addition, the model assumes that the adsorption process does not follow a strictly Langmuir or Freundlich pattern but a combination of both. This assumption may not hold for all adsorption systems, especially those that strictly adhere to either Langmuir or Freundlich behavior. Furthermore, the Redlich-Peterson model lacks a clearly established physical basis and is thus an empirical model similar to the Sips model [70]. Hence, we will not offer a mechanical interpretation based on the best model because it is a variation of the Sips equation, which is an empirical model in and of itself.

Fritz-Schlunder III

The Fritz Schlunder III isotherm serves as a model utilized in explaining adsorption processes, on surfaces with differing properties. It blends aspects of the Langmuir and Freundlich isotherms making it applicable to scenarios where adsorption sites exhibit variations in both energy levels and capacities. This model proves to be highly effective in real world data for adsorption systems like organic pollutant adsorption on activated carbon or heavy metal removal from wastewater using biosorbents. Its capability to accommodate adsorption energies and capacities establishes it as a tool, for predicting adsorption tendencies within heterogeneous systems.

Table 3. Isothermal models' constants for the top seven models.

| Model | | Unit | Value | (95% confidence in- terval) |
|----------------|--------------|--------------------|-------|--------------------------------|
| Unilan | q_{mU} | mg g ⁻¹ | 16.79 | -5.873 to 39.445 |
| | b_U | | 2.35 | 0.518 to 4.178 |
| | a_U | L mg ⁻¹ | 0.08 | 0.022 to 0.129 |
| Redlich-Peter- | K_{RPI} | L g ¹ | 12.91 | 4.412 to 21.405 |
| son | K_{RP2} | $(mg L^{-1})^{-n}$ | 0.27 | -0.044 to 0.580 |
| | β_{RP} | | 0.89 | 0.779 to 0.999 |
| Moreau | q_{mM} | mg g ⁻¹ | 90.82 | 76.336 to 105.303 |
| | b | L mg ⁻¹ | 0.15 | 0.089 to 0.203 -0.049 |
| | l | - | 0.3 | to 0.646 |
| Fritz-Schlun- | q_{mFS} | mg g ⁻¹ | 48.24 | 22.653 to 73.816 |
| der III | n_{FS} | | 0.89 | 0.779 to 0.999 |
| | K_{FS} | L mg ⁻¹ | 0.27 | -0.044 to 0.580 |
| Toth | q_{mS} | mg g ⁻¹ | 45.93 | 20.956 to 70.912 |
| | K_T | $(mg L^{-1})^{nT}$ | 4.10 | -0.322 to 8.514 |
| | n_T | | 0.94 | 0.879 to 0.998 |
| Langmuir | q_{mL} | mg g ⁻¹ | 81.93 | 76.422 to 87.440 |
| - | b_L | L mg ⁻¹ | 0.10 | 0.070 to 0.133 |

A comparison of the SDS adsorption capacity by various adsorbents reported in the literature is presented in Table 4. The types of regression used in the study are also displayed and most of the studies used linear regression to analyse their data. According to Paranjape and Sadgir [71], linear regression is the most frequently used method to determine the best-fitting model for adsorption isotherms and kinetics, including the respective model

parameters [5,72–75]. However, the error distribution can vary significantly depending on the linearization of the isotherm and kinetic equations, leading to either the best or worst fit. Therefore, many researchers prefer the nonlinear regression approach to verify the parameters of adsorption isotherms and kinetics. This method is based on minimizing the discrepancies between the experimental data and the predicted isotherm values.

Table 4. Summary of SDS sorption by various adsorbents.

| Adsorbents | pН | Adsor- bent dosage (g/L) | Maximum Adsorption capacity O _{max} (mg/g) | Isotherm models | Kinetic models | e Regres- s sion | Error func- tion | Ref |
|--|-----|-----------------------------------|--|---------------------------------|-------------------|---------------------|------------------------|------|
| Chemically activated co- conut shell | - | 2 | 90.9 | Langmuir | PSO | Linear | <i>R</i> ² | [5] |
| Medicinal plants | 5 | 25 | - | Freundlich | PFO | Linear | R^2 | [72] |
| Sweet lime peel charcoal | 4.3 | 2 g | - | Langmuir | n.a. | Linear | R^2 | [73] |
| Granular acti- vated char- coal | 7 | 5 | 3.750 | Langmuir | n.a. | Linear | R^2 | [74] |
| Waste tire rubber gran- ules | 7 | 5 | 4.164 | Langmuir | n.a. | Linear | R^2 | [74] |
| Wood char- coal | 7 | 5 | 5.170 | Freundlich | n.a. | Linear | R^2 | [74] |
| Silica gel | 7 | 5 | 5.181 | Freundlich | n.a. | Linear | R^2 | [74] |
| Modified cel- lulose | 7 | n.a | 32.5 | Langmuir | PSO | Linear | R^2 | [75] |
| Alumina | 4 | n.a | 230 | 2-step ad- sorption model | n.a | Non-lin- ear | n.a | [76] |

n.a. PSO Not available

Pseudo second order

Pseudo first order PFO Coefficient of determination

CONCLUSION

Using a variety of models with one to three parameters each, the adsorption isotherm data for SDS on activated coconut shell was subjected to nonlinear regression. Moreau model outperformed Unilan, Redlich-Peterson, Fritz-Schlunder III, Toth, and Langmuir, in that order, according to statistical analysis based on error function assessments. Results from experiments were more consistent with the upper limits of adsorption capacities predicted by the Moreau and Langmuir models. The value of the maximum monolayer adsorption capacity for SDS binding to activated coconut shell according to the Langmuir's parameter q_{mL} was 81.93 mg g⁻¹ (95% Confidence interval from 76.422 to 87.440), while b_L (L mg⁻¹), the Langmuir model constants was 0.10 L mg⁻¹ (95% C.I. from 0.070 to 0.133). The value of the maximum monolayer adsorption capacity for SDS binding to activated coconut shell according to the Moreau's parameter q_{mM} was 90.82 mg g⁻¹ (95%) Confidence interval from 76.336 to 105.303), while b (L mg⁻¹), the Moreau's model constant was 0.15 L mg⁻¹ (95% C.I. from 0.089 to 0.203) and l, another Moreau's dimensionless constant was 0.3 (95% C.I. from -0.049 to 0.646). These values include 95% confidence intervals, which were absent in the original work.

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