

JOURNAL OF ENVIRONMENTAL BIOREMEDIATION AND TOXICOLOGY



Website: http://journal.hibiscuspublisher.com/index.php/JEBAT/index

Normality, Grubb's and Runs Tests for the von Bertalanffy Model Used in the Fitting of the Growth of *Bacillus cereus* strain wwcp1 on Malachite Green Dye

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HISTORY

Received: 4th April 2022 Received in revised form: 15th July 2022 Accepted: 25th July 2022

KEYWORDS

Wald–Wolfowitz runs test Residual von Bertalanffy model outlier Grubb's test

ABSTRACT

When diagnostic tests show that the residuals form a pattern, there are a few treatment alternatives to choose from. Two of these choices are running a nonparametric analysis and switching to a different model. In this study, the Wald-Wolfowitz runs test is utilized as a statistical diagnosis approach to determine whether or not the randomization criteria have been met. The Wald-Wolfowitz runs test was chosen as the best method to use for this research since it was necessary to examine the randomness of the residual for the von Bertalanffy model used in the fitting of the growth of *Bacillus cereus* strain wwcp1 on malachite green dye. The observations indicated that the residual series had an adequate number of runs; this was the outcome. The runs test discovered four runs, despite the fact that the randomization assumption predicted thirteen runs. This implies that the runs in the residual collection are only marginally meaningful. The fact that the p-value was less than 0.05 indicates that the null hypothesis is rejected; this implies that the residuals include convincing evidence of non-randomness. Furthermore, this demonstrates the need of looking at potential outliers. However, the test for the lack of an outlier does not demand reanalysis of the data, as indicated by Grubb's test findings. To resolve this discrepancy, either a different model needs to be used or more data needs to be added.

INTRODUCTION

A biological system can be viewed as a collection of different cellular compartments (such as cell types), each of which is specialized for a certain biological purpose (e.g. white and red blood cells have very different commitments). An object is a type of elemental unit that can be investigated but whose internal structure is unknown or non-existent. The scale used to depict the system will be decided by the elemental unit chosen. The availability of data covering a wide range of biological states and processes, as well as the temporal interdependence of those activities, allows for biological system study at a variety of organizational levels. These stages start at the molecular level and work their way up to the population level. A model is a generalized representation of a system that researchers can decode or interpret. Models are widely used in computer simulations. A model is a description of a system that focuses on its components and the interactions that occur between them. The availability of data covering a wide range of biological states and processes, as well as the temporal interdependence of those activities, allows for biological system study at a variety of organizational levels. These stages start at the molecular level and work their way up to the population level. In high-throughput genomics and proteomics research, the use of mathematical and computer models to aid in the interpretation of biological data is becoming more common.

The later stages of the research process, hypothesis creation and experiment prescription, are both facilitated by the use of complicated computer models that allow for the depiction of intricate biological processes. Computational models are now using knowledge discovery approaches to make advantage of the huge amounts of data collected in biomedical databases. This is done to make the best use of the given information. The availability of data covering a wide range of biological states and processes, as well as the temporal interdependence of those activities, allows for biological system study at a variety of organizational levels. These stages start at the molecular level and work their way up to the population level. In many circumstances, the observed phenomenon's relationship to time or concentration can be statistically represented using least square techniques, which are often employed in nonlinear regression [1–4].

In any case, the residuals of a nonlinear regression must have natural dispersion. This contrasts from the usual least squares approach, which requires linear regression residuals to have a normal distribution. This is because the concept of least squares is the foundation of the traditional least square's technique. The far more important residuals must be random and have the same variance (homoscedastic distribution). The Wald-Wolfowitz runs test can be used to determine the effectiveness of the randomization approach (Motulsky and Ransnas 1987). In order for the usual least squares technique to work effectively, the residuals of the curve in a nonlinear regression must have a natural dispersion, whereas in a linear regression, the residues must have a normal distribution. The residuals must be randomly distributed and have the same variance (homoscedastic distribution) (homoscedastic distribution). The Wald-Wolfowitz runs test is used to determine if the trial was randomized or not. In this study the von Bertalanffy model used in the fitting of the growth of Bacillus cereus strain wwcp1 on malachite green dye was assessed for its non-randomness of residuals and the potential presence of an outlier.

METHODOLOGY

Residual data

One of the benefits of residual information is that it may be used to assess the correctness of any model when fitting a curve in nonlinear regression. (D'Agostino, 1986). In statistics, residual data is estimated as the difference between observed and anticipated data, the latter generated using an appropriate model and typically performed using nonlinear regression. (Eqn. 1);

$$e_i = y_i - f\left(x_i; \hat{\beta}\right) \tag{1}$$

where y_i is the *i*th response from a particular data and x_i is the vector of descriptive variables to each set at the *i*th observation which corresponds to values from a particular data. Residual data from in this study, the von Bertalanffy model used in the fitting of the growth of *Bacillus cereus* strain wwcp1 on malachite green dye was assessed from a previous work [5].

Grubbs' Statistic

The test is a statistical technique for identifying outliers in a univariate data set with a Gaussian or normal distribution. Grubb's test assumes that the data is spread uniformly. In a univariate context, the test is used to find outliers [6]. The test can be utilized to the maximal or minimal examined data from a Student's t distribution (Eq. 2) and to test for both data instantaneously (Eq. 3).

$$G_{\min} = \frac{X - \min(X)}{s}$$

$$G_{\max} = \frac{\max(X) - \bar{X}}{s}$$

$$p_{G} = 2n \cdot p_{t} \left(G \frac{\sqrt{n(n-2)}}{n-1}, n-2, 1 \right)$$

$$G_{all} = \frac{\max(\bar{X} - \min(X), \max(X) - \bar{X})}{s}$$

$$p_{G} = n \cdot p_{t} \left(G \frac{\sqrt{n(n-2)}}{n-1}, n-2, 2 \right)$$
(3)

The ROUT method can be employed in the event that there is more than one outliers [7]. The False Discovery Rate is the foundation of the approach (FDR). Q, a probability of (incorrectly) recognizing one or more outliers must be explicitly specified. It is the highest desired FDR. Q is fairly comparable to alpha in the absence of outliers. Assumption that all data has a Gaussian distribution is mandatory.

Runs test

When doing a nonlinear regression, the residuals of the curve must have a natural distribution. This is in contrast to the least squares approach, which requires that the residues have a normal distribution. These specs are available here. Furthermore, residuals must be random and have the same variance (homoscedastic distribution). The Wald-Wolfowitz test is used to assess whether or not randomization has been completed. The concept is statistically valid because biological systems are inherently unpredictable [8–10]. This test was run on the regression residuals to detect unpredictability.

The number of sign runs is sometimes expressed as a percentage of the maximum number possible. The runs test looks at a sequence of residuals that include both positive and negative values. Following the test, a good run is frequently described by the existence of an alternating or adequately balanced number of positive and negative residual data. The runs test determines if the residuals data contains an excessive or inadequate amount of runs of sign (**Eq. 4**). Too few runs may indicate systematic bias or a clustering of residuals with the same sign, but too many with the same run sign may indicate negative serial correlation [11,12]. The test statistic is

 $H_0=$ the sequence was produced randomly $H_a=$ the sequence was not produced randomly

$$Z = \frac{R - R}{sR} \tag{4}$$

Where Z is the test statistic, \overline{R} indicates the anticipated number of runs, sR is the standard deviation of the runs and R is the observed number of runs and (Eqs. 5 and 6). The calculation of the respective values of R and sR (n_1 is positive while n_2 is negative signs) is as follows.

$$\bar{R} = \frac{2n_1 \cdot n_2}{n_1 + n_2} + 1$$

$$s^2 R = \frac{2n_1 \cdot n_2 (2n_1 \cdot n_2 - n_1 - n_2)}{(n_1 + n_2)^2 (n_1 + n_2 - 1)}$$
(6)

As an example Test statistic: Z = 3.0

Significance level: $\alpha = 0.05$

Critical value (upper tail): $Z_{1-\alpha/2} = 1.96$

Critical region: Reject H₀ if |Z| > 1.96

If the test statistical value (Z) is greater than the critical value, then the rejection of the null hypothesis at the 0.05 significance level shows that the sequence was not generated randomly.

RESULTS AND DISCUSSION

Finding a good model for biological and even chemical processes may be difficult. The modelling approach is complex on its own, and mistakes are common. The modelling technique is a procedure in and of itself that adheres to a set of loosely specified criteria. Four essential actions are required to complete the technique. The first step is to thoroughly comprehend the situation at hand, which requires precisely stating the model's questions. The second step is developing a problem-solving strategy, which includes outlining a set of actions that must be done in order to locate an accurate model of the system under examination. In this step, you will collect knowledge and data from experts in the field as well as published works, model structure, model hypothesis, conceptual model, appropriate mathematical formalism selection, solving the formal model, obtaining results, determining whether the model's results match the data available, and other similar tasks. The third part is to put the plan into action, which involves repeating the methods from the previous two stages, determining whether or not the solution is right, and finally enhancing the model. This final phase is a critical test for validating the hypothesis developed prior to the model's setup. Finally, all models must be mathematically fit, which is where nonlinear regression comes in.

The residuals data are crucial in the statistical analysis of nonlinear regression, which is performed using nonlinear regression. The residuals show the discrepancy between the data collected and the data expected to be collected. The gap between the values predicted by a mathematical model and the values observed in the data is referred to as "residuals." These discrepancies may be seen when comparing the model's predictions to the data. The residuals must be statistically analyzed to ensure that they are sufficiently random, lack outliers, have a normal distribution, and do not exhibit autocorrelation. This may be done by determining whether or not they have any outliers. It is common practice to show residue data as positive and negative values. This is a key component for assuring data balance, and it is visible before any tests are run. It is common practice to reject the results of any residual tests when employing nonlinear regression. In general, the quality of a model is considered weaker when there is a greater difference between anticipated and observed values. This is because the two sets of data have a lower degree of correlation, which explains why this is the case [13]. The residuals for the von Bertalanffy model are shown in Table 1.

Table 1. Residuals for the von Bertalanffy model used in the fitting o	f
the growth of Bacillus cereus strain wwcp1 on malachite green dye.	

Time	Residuals
0	-0.3
10	0.4
18	0.17
30	-0.19
39	-0.23
49	-0.25
59	-0.26
69	-0.2
79	-0.12
90	-0.09
100	-0.08
110	-0.03
120	0.02
129	0.05
140	0.05
150	0.06
159	0.07
169	0.09
180	0.1
189	0.1
199	0.11
209	0.12
219	0.13
229	0.13
239	0.13

Grubbs' test indicated no indication of an outlier when applied to previously available data. This demonstrates that the model correctly described the data. When attempting to fit a nonlinear curve, it is easy to introduce large errors if either the mean or a single data point from a triple is twisted. Both of these circumstances have the same outcome. The Grubbs test can identify a single abnormality throughout any given time period. It is critical to seek for and delete any outlying data points when fitting curves [2,14–19]. Because this data point was judged an outlier, it was eliminated from the collection, and the analysis was repeated until no further outliers were found. Sample sizes of six or below are not recommended since the test consistently flags the vast majority of points as outliers. Furthermore, repeating the test may change the likelihood of it discovering something.

The Grubbs' test statistic zeroes in on the sample value with the largest absolute deviation from the sample mean, as indicated by the sample's standard deviation, to select the winner. If the result of the test statistic g is larger than the critical value, the result is termed an outlier. This is due to the fact that the critical value is the lowest value that can be tolerated. Grubbs' test results revealed the absence of outlier (Table 2). An extreme data point that the investigator thinks implausible because it lacks a number of certain qualities is an example of a possible outlier. An outlier is a figure that stands out as being notably different from the bulk of the other data in a sample. An outlier is, for example, a maximum that is statistically significantly bigger than the maximum distribution predicted by the population model employed in engineering. Using these criteria, one may assess if the highest value is an anomaly.

The Chauvenet's criteria, the 3-sigma criterion, and the Zscore are all approaches for determining if a measurement contains likely outliers. The 3-sigma criteria and the Z-score are two popular statistical methods in the field of chemometrics. A boxplot may be used to rapidly and readily identify outliers in measurement data. Despite the fact that the methods in question are simple, quick, and pass visual inspections, it is recommended that a statistical test be performed to establish whether or not a data set contains an outlier. Dixon's Q-test and Grubbs' ESD-test are two specific tests that may be used to assess whether or not a

person is an outlier. Grubbs created both of these examinations [20].

 Table 2. Outlier detection from the residuals of the von Bertalanffy

 model used in the fitting of the growth of *Bacillus cereus* strain wwcp1

 on malachite green dye.

row	Value	Z	Significant Outlier?
1	-0.3	1.7685	
			Furthest from the rest, but not a
2	0.4	2.369	significant outlier ($P > 0.05$).
3	0.17	1.0095	c ,
4	-0.19	1.1183	
5	-0.23	1.3547	
6	-0.25	1.4729	
7	-0.26	1.532	
8	-0.2	1.1774	
9	-0.12	0.7045	
10	-0.09	0.5272	
11	-0.08	0.4681	
12	-0.03	0.1726	
13	0.02	0.1229	
14	0.05	0.3003	
15	0.05	0.3003	
16	0.06	0.3594	
17	0.07	0.4185	
18	0.09	0.5367	
19	0.1	0.5958	
20	0.1	0.5958	
21	0.11	0.6549	
22	0.12	0.714	
23	0.13	0.7731	
24	0.13	0.7731	
25	0.13	0.7731	

Before the Grubbs test findings to be deemed valid, the specific value of the anticipated number of outliers, denoted by the letter k, must be specified. This is by far the most significant constraint imposed by the exam. It is very possible that the exam results will be tainted if k is not appropriately reflected in the test. This is not an unthinkable prospect. In circumstances when there are several outliers or the exact number of outliers cannot be identified, the Rosner's generalised Extreme Studentized Deviate test, often known as the ESD-test or the ROUT approach, might be used. The Extreme Studentized Deviation exam refers to both of these assessments. In common terminology, both of these procedures are known to as the ESD-test [7] are recommended [21]. The ROUT approach, which combines robust regression and outlier removal, is becoming more popular in the elimination of numerous outliers [14,22–25]

The runs test revealed that there were 4 total runs, whereas the randomization assumption predicted 13 runs (**Table 3**). This implies that the runs included in the residual collection are just those that are marginally important. The Z-value is the number of standard errors by which the actual number of runs differs from the predicted number of runs, and the p-value indicates how important this Z-value is in terms of the difference between the two values. It should be evaluated similarly to any other statistic that employs p-values. If the p-value is less than 0.05, which is the cutoff point at which the null hypothesis may be rejected, it is possible to conclude that the residuals are not fully random. When the threshold is reached, this is the situation.

The fact that the p-value is smaller than 0.05 implies that the null hypothesis is rejected; this demonstrates that there is convincing evidence of the residuals' non-randomness. When there are a large number of occurrences of a particular run sign, this may indicate a negative serial correlation; when there are a small number of runs, this may indicate a clustering of residuals with the same sign or the presence of a systematic bias; and when there are a large number of occurrences of a particular run sign, this may indicate a positive serial correlation; and when there are a large number of occurrences of a particular run sign, this may indicate a positive serial correlation; and when there are

a large number of occurrences of a particular run sign, this may indicate [12]. The runs test can discover a systematic deviation from the curve, such as an overestimation or underestimating of the sections, when a specific model is applied. This is accomplished by contrasting the actual and anticipated values.

Table 3. Runs test for randomness.

Runs test	Residual data set
R=	4
n0=	10
n1=	15
n=	25
E(R)=	13.00
Var(R)=	5.50
StDev(R)=	2.35
Z=	-3.84
p-value=	0.0001
Note: E(R) Expected run	1

This may be performed by comparing the model's predictions to the actual outcomes. The runs test is used to detect if there are an excessive number of signs runs and whether there are insufficient runs in general. A runs test was performed on the regression residuals to assess whether or not there was evidence of nonrandomness. It is possible to build a model with an ordered variance of the curve that is more or less than the estimate. This is only one of the ways that something like this is possible. As part of the run test, which is used to establish whether or not a chemical poses a risk to human health, a comparison is done between a drug's normally negative sequence of residues and its typically positive sequence.

This test is used to determine whether a substance is hazardous to human health. A notable occurrence is usually differentiated from other conceivable outcomes by a change or combination of shifts between negative and positive residual values. The transition from one condition to another, or a succession of changes, is frequently what characterizes a remarkable conclusion [11]. The use of the largest number of indicators that may be counted is a frequent practice in this discipline. The run's test is used to determine whether a large number of signs passes, or a small number of sign passes are more likely. Run signs may imply negative serial correlation, but it is also possible that residues are connected with the same sign or that systemic biases are influencing the results [12].

When testing time-series regression models to identify the existence or absence of autocorrelation, it is usual practise to utilise the run technique. According to the conclusions of study that used Monte Carlo simulation, run-time testing is responsible for unequal error rates in both tails of the distribution. This study implies that run-time autocorrelation research may be unstable, and it predicts that in the not-too-distant future, the Durbin-Watson methodology will be the most often used method for analysing autocorrelation [26]. It has been demonstrated that the approach utilised in this analysis, which is based on earlier research on the unpredictability of residuals, is reliable. For example, modelling the growth curve of algae using the Baranyi-Roberts model, which provides statistical sufficiency [27], Moraxella sp. B on monobromoacetic acid (MBA) [9] and the Buchanan-three-phase model used in the fitting the growth of Paracoccus sp. SKG on acetonitrile [28]. For lead (II) absorption by alginate gel bead, the runs tests on the residuals for the Sips and Freundlich models were found to be sufficient [29].

It has been shown that the strategy used in this analysis, which was derived from previous research on the unpredictability of residuals, is dependable. One can anticipate the growth curve of algae using the Baranyi-Roberts model, which assesses whether or not statistical sufficiency has been fulfilled [30]. In the body of academic research, different applications of the runs test of residual may be found for the purpose of evaluating the validity of the nonlinear regression [31-35]. However, the test for the lack of an outlier does not demand reanalysis of the data, as indicated by Grubb's test findings. To resolve this discrepancy, either a different model needs to be used or more data needs to be added.

CONCLUSION

The Wald-Wolfowitz runs test was performed as part of the scope of this investigation. The observations revealed that the residual series had an acceptable number of runs; this was the result of the observations. The runs test found a total of four runs, although the randomization assumption expected thirteen runs. This means that the runs included in the residual collection are only slightly significant. The fact that the p-value is less than 0.05 suggests that the null hypothesis is rejected; this demonstrates that there is compelling evidence of non-randomness in the residuals. Furthermore, this illustrates the need of investigating potential outliers. This illustrates, in particular, that extra intervention is required to detect any outliers that may exist in the data. The results of Grubb's test suggest, however, that testing for the absence of an outlier does not necessitate reanalyzing the data. Either a different model or additional data is required to explain the discrepancy.

REFERENCES

- Lee KB, Lee JM, Park TS, Lee SH. Construction of classical 1. confidence regions of model parameters in nonlinear regression analyses. Appl Radiat Isot. 2010 Jul 1;68(7):1261-5.
- Fong Y, Wakefield J, De R, Frahm N. A Robust Bayesian Random 2 Effects Model for Nonlinear Calibration Problems. Biometrics. 2012;68(4):1103-12.
- 3. Huang L. Direct construction of predictive models for describing growth of Salmonella Enteritidis in liquid eggs - A one-step approach. Food Control. 2015;57:76-81.
- Yun B, Yu X, Xue M, Liu Y, Wang J, Wu X, et al. Effects of dietary 4. protein levels on the long-term growth response and fitting growth models of gibel carp (Carassius auratus gibelio). Anim Nutr [Internet]. 2015 May 31 [cited 2015 Jun 1]; Available from: http://www.sciencedirect.com/science/article/pii/S2405654515000 268
- 5. Yahuza S, Sabo IA. Mathematical Modelling of the Growth of Bacillus cereus Strain wwcp1on Malachite Green Dye. J Biochem Microbiol Biotechnol. 2021 Dec 31;9(2):25-9.
- Grubbs F. Procedures for detecting outlying observations in 6. samples. Technometrics. 1969;11(1):1-21.
- Motulsky HJ, Brown RE. Detecting outliers when fitting data with 7. nonlinear regression - A new method based on robust nonlinear regression and the false discovery rate. BMC Bioinformatics. 2006;7.
- Lazaridis NK, Asouhidou DD. Kinetics of sorptive removal of 8. chromium(VI) from aqueous solutions by calcined Mg-Al-CO3 hydrotalcite. Water Res. 2003 Jul 1;37(12):2875-82.
- Sabullah MK, Shukor MS, Masdor NA, Shamaan NA, Shukor MY. Test of randomness of residuals for the Buchanan-three-phase model used in the fitting the growth of Moraxella sp. B on monobromoacetic acid (MBA). Bull Environ Sci Manag. 2015;3(1):13-5.
- Uba G, Mansur A, Manogaran M, Shukor MY. Runs Test for the 10. Residuals of The Morgan-Mercer-Flodin (MMF) Model Used for Modelling the Total Number of Covid-19 Cases for Brazil. Bull Environ Sci Sustain Manag. 2021 Jul 31;5(1):7-11.
- 11. Motulsky HJ, Ransnas LA. Fitting curves to data using nonlinear regression: a practical and nonmathematical review. FASEB J Off Publ Fed Am Soc Exp Biol. 1987;1(5):365-74.
- 12 Draper NR, Smith H. Applied Regression Analysis. Wiley, New York; 1981.

- 13. López S, Prieto M, Dijkstra J, Dhanoa MS, France J. Statistical evaluation of mathematical models for microbial growth. Int J Food Microbiol. 2004;96(3):289-300.
- 14. Tietjen GL, Moore RH. Some Grubbs-Type Statistics for the Detection of Several Outliers. Technometrics. 1972;14(3):583-97.
- Bendre SM, Kale BK. Masking effect on tests for outliers in normal 15. samples. Biometrika. 1987 Dec 1;74(4):891-6.
- Barnett V, Lewis T. Outliers in Statistical Data. 3rd ed. Chichester; 16. New York: Wiley; 1994. 604 p.
- Jain RB. A recursive version of Grubbs' test for detecting multiple 17. outliers in environmental and chemical data. Clin Biochem. 2010:43(12):1030-3
- 18. Cohn TA, England JF, Berenbrock CE, Mason RR, Stedinger JR, Lamontagne JR. A generalized Grubbs-Beck test statistic for detecting multiple potentially influential low outliers in flood series. Water Resour Res. 2013;49(8):5047-58.
- 19 Miller JN. Using the Grubbs and Cochran tests to identify outliers. Anal Methods. 2015;7(19):7948-50.
- 20 Barnett V, Lewis T. Outliers in Statistical Data. 3rd ed. Chichester ; New York: Wiley; 1994. 604 p.
- Rosner B. Fundamentals of biostatistics. 7th ed. Boston: 21 Brooks/Cole: 2011.
- 22. Horn R, Stewart AJ, Jackson KV, Dryburgh EL, Medina-Torres CE, Bertin FR. Clinical implications of using adrenocorticotropic hormone diagnostic cutoffs or reference intervals to diagnose pituitary pars intermedia dysfunction in mature horses. J Vet Intern Med. 2021;35(1):560-70.
- 23. Shukor MY. Outlier and Normality Testing of the Residuals from the Carreau-Yasuda Model in Fitting the Rheological Behavior of the Non-Newtonian fluid TF2N. Bioremediation Sci Technol Res. 2021 Jul 31;9(1):20-6.
- 24. Uba G, Zandam ND, Mansur A, Shukor MY. Outlier and Normality Testing of the Residuals for the Morgan-Mercer-Flodin (MMF) Model Used for Modelling the Total Number of COVID-19 Cases for Brazil. Bioremediation Sci Technol Res. 2021 Jul 31;9(1):13-9.
- Li J. A Practical \$O(N^2)\$O(N2) Outlier Removal Method for 25. Correspondence-Based Point Cloud Registration. IEEE Trans Pattern Anal Mach Intell. 2022 Aug;44(8):3926-39.
- 26. Huitema BE, McKean JW, Zhao J. The runs test for autocorrelated errors: unacceptable properties. J Educ Behav Stat 1996;21(4):390-404.
- 27. Halmi MIE, Shukor MS, Johari WLW, Shukor MY. Evaluation of several mathematical models for fitting the growth of the algae Dunaliella tertiolecta. Asian J Plant Biol. 2014;2(1):1-6.
- 28. Gunasekaran B, Shukor MS, Masdor NA, Shamaan NA, Shukor MY. Test of randomness of residuals for the Buchanan-three-phase model used in the fitting the growth of Paracoccus sp. SKG on acetonitrile. J Environ Bioremediation Toxicol. 2015;3(1):12-4.
- 29. Cataldo S, Gianguzza A, Merli M, Muratore N, Piazzese D, Turco Liveri ML. Experimental and robust modeling approach for lead(II) uptake by alginate gel beads: Influence of the ionic strength and medium composition. J Colloid Interface Sci. 2014 Nov 15;434:77-
- 30. Umar G. Test of the Randomness of Residuals for the Pseudo-1st Order Kinetic Modelling of Adsorption of the Brominated Flame Retardant 4-bromodiphenyl Ether onto Biochar-immobilized Sphingomonas sp. Bioremediation Sci Technol Res. 2021 Jul 31;9(1):27-31.
- 31. Cooper SM, Baker JS, Eaton ZE, Matthews N. A simple multistage field test for the prediction of anaerobic capacity in female games players. Br J Sports Med. 2004;38(6):784-9.
- Worthington AC, Higgs H. Efficiency in the Australian stock 32. market, 1875-2006: A note on extreme long-run random walk behaviour. Appl Econ Lett. 2009;16(3):301-6.
- 33. Abu GA, Abachi PT, Oloja-Ojabo ED. Long-run relationship between agricultural crop prices and supply response in Benue State, Nigeria: 1990-2010. Eur J Soc Sci. 2011;24(4):565-75.
- 34. Burns RD, Hannon JC, Brusseau TA, Eisenman PA, Shultz BB, Saint-Maurice PF, et al. Development of an aerobic capacity prediction model from one-mile run/walk performance in adolescents aged 13-16 years. J Sports Sci. 2016;34(1):18-26.
- Gardiner SK, Mansberger SL. Effect of restricting perimetry testing 35. algorithms to reliable sensitivities on test-retest variability. Invest Ophthalmol Vis Sci. 2016;57(13):5631-6.