

# BIOREMEDIATION SCIENCE AND TECHNOLOGY RESEARCH

Website: https://journal.hibiscuspublisher.com/index.php/BSTR



## Modelling the Effect of Heavy Metal on the Growth Rate of an SDSdegrading *Pseudomonas* sp. strain DRY15 from Antarctic soil

Motharasan Manogaran<sup>1</sup>, Ahmad Razi Othman<sup>2</sup>, Mohd Yunus Shukor<sup>1</sup> and Mohd Izuan Effendi Halmi<sup>3</sup>\*

<sup>1</sup>Department of Biochemistry, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia,

43400 UPM Serdang, Selangor, D.E, Malaysia.

<sup>2</sup>Department of Chemical Engineering, Faculty of Engineering and Built Environment, Universiti Kebangsaan Malaysia,

43600 UKM Bangi, Selangor, D.E, Malaysia.

<sup>3</sup>Department of Soil Management, Faculty of Agriculture, Universiti Putra Malaysia,

43400 UPM Serdang, Selangor, D.E, Malaysia.

\*Corresponding author: Dr. Mohd Izuan Effendi Halmi Department of Soil Management, Faculty of Agriculture, Universiti Putra Malaysia, 43400 UPM Serdang, Selangor, D.E, Malaysia.

Email: m\_izuaneffendi@upm.edu.my

### HISTORY

Received: 4<sup>th</sup> April 2019 Received in revised form: 7<sup>th</sup> of June 2019 Accepted: 24<sup>th</sup> of July 2019

KEYWORDS

SDS-degrading Pseudomonas Antarctica modified Gompertz Shukor model

## ABSTRACT

The SDS-degrading bacterium Pseudomonas sp. strain DRY15 was strongly inhibited by heavy metals especially mercury. Growth of the SDS-degrading bacterium at various concentrations of mercury shows a sigmoidal pattern with lag periods ranging from 7 to 10 h. As the concentration of mercury was increased, the overall growth was inhibited with 1.0 g/L causing an almost cessation of bacterial growth. The modified Gompertz model was utilized to obtain growth rates at different concentrations of mercury. The growth rates obtained from the modified Gompertz model was then modelled according to the modified Han-Levenspiel, Wang, Liu, modified Andrews, the Amor and Shukor models. Out of the five models, only the Shukor, Wang, modified Han-Levenspiel and the Liu models were able to fit the curve, whilst the modified Andrews and Amor models were unable to fit the curves. The best model was Shukor based on the lowest values for RMSE and AICc, highest adjusted correlation coefficient ( $adR^2$ ) and values of AF and BF closest to unity. The parameters obtained from the Shukor model, which are  $S_m$ ,  $\mu_{max}$  and n which represent critical heavy metal ion concentration (mg/L), maximum growth rate  $(h^{-1})$  and empirical constant values were 6.0 (95%, confidence interval from 5.87 to 6.14), 0.09 (95%, confidence interval of 0.086 to 0.096) and 4.2 (95%, confidence interval from 3.1 to 5.2), respectively.

## INTRODUCTION

The setting up of many bases and the increase in the numbers of tourists and tourist's ships in the cold region has introduced anthropogenic pollutants in Antarctica. Anionic surfactant such as sodium dodecyl sulphate has been reported to occur in the Antarctic Maxwell Bay and its adjacent sea areas at concentrations of up to 1.0 mg l<sup>-1</sup> [1] and biodegradation of the surfactant by sea water bacteria has been reported [2]. Detergents are known to have detrimental effects to aquatic life [3–5]. According to literature data, anionic surfactants give toxic effects to various aquatic organisms at concentrations from 0.0025 to 300 mg l<sup>-1</sup> [6]. Another study indicated that exposure to SDS has a detrimental effect on oyster digestive gland, resulting in the perturbation of the metabolic and nutritional functions, and

having a direct influence on oyster survival [7]. Toxicity towards invertebrates and crustaceans could occur as the considerable number of anionic surfactants released into water. The life cycle of aquatic animals has also been influenced by the anionic surfactants, modified the behavior of the fish such as erratic movements, muscle spasms and body torsion [8]. Due to this, remediation of SDS is of vital importance.

Microbes are known for their ability to degrade organics including SDS [9–15], and their use as bioremediation agents is important for economical removal of xenobiotic pollutants. Biodegradation of anionic surfactant under aerobic conditions by the bacterium *Pseudomonas* sp. strain C12B was among the first to be studied [16], and to date quite a number of SDS-degrading bacteria have been isolated and characterized [10,11,17–29].

Works on cold-adapted microbes with ability to degrade SDS are rare [11,30]. We have previously reported the isolation of such a bacterium and found that the degradation of detergent is strongly affected by heavy metals-zinc especially. In this work we model the inhibitory effect of SDS on the growth rate of this bacterium using several models.

### MATERIALS AND METHODS

#### Growth and maintenance of bacterium

The bacterium was sourced from our inhouse culture collection unit. The bacterium was grown and maintained on an SDS enrichment media in a 250 ml conical flask, incubated at 15 °C with shaking at 150 rpm on an orbital shaker. The basal salts (BS) medium contained the followings: KH<sub>2</sub>PO<sub>4</sub>, (1.36 g l<sup>-1</sup>), Na<sub>2</sub>HPO<sub>4</sub>, (1.39 g L<sup>-1</sup>), KNO<sub>3</sub>, (0.5 g L<sup>-1</sup>), MgSO<sub>4</sub> (0.01 g L<sup>-1</sup>),  $CaCl_2$  (0.01 g L<sup>-1</sup>) and (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> (7.7 g L<sup>-1</sup>). The medium also contained the following trace elements: ZnSO4.7H2O (0.01 g L <sup>1</sup>), MnCl<sub>2</sub>.4H<sub>2</sub>O (0.01 g L<sup>-1</sup>), H<sub>3</sub>BO<sub>4</sub> (0.01 g L<sup>-1</sup>), CoCl<sub>2</sub>.6H<sub>2</sub>O (0.01 g L<sup>-1</sup>), FeSO<sub>4</sub>.2H<sub>2</sub>O (0.01 g L<sup>-1</sup>), CuCl<sub>2</sub>.2H<sub>2</sub>O (0.01 g L<sup>-1</sup>) and Na2MoO4.2H2O (0.01 g L-1) [18]. Filter-sterilized sodium dodecyl sulphate was added into the medium as a carbon source at the final concentration of 1.0 g l-1. Maintenance of the bacterium every fortnight was carried out on agar plate (1.5% agar) supplemented with solid sodium dodecyl sulphate. The growth rate of the bacterium at 0.1% SDS (w/v) in the presence of various concentrations of zinc of up to 6 mg  $L^{-1}$  was monitored under the same optimum conditions previously reported [12].

#### Primary growth modelling on SDS

The maximum specific growth rate on SDS was modelled according to the modified Gompertz model as this model is routinely used in modelling the growth of microorganisms on xenobiotics [31–37]. The equation (Eqn. 1) is as follows;

$$y = A \exp\left\{-\exp\left[\frac{\mu_m e}{A}(\lambda - t) + 1\right]\right\}$$
 (Eqn. 1)

The value obtained from this primary modelling exercise was then used to model the effect of metal as follows;

#### Effect of metal on growth rate of on SDS

The models utilized in this study is as follows (Table 1);

Table 1. Metal inhibition models.

Models	Equation	Ref	
Modified Han- Levenspiel	$r = u_{max} \left( 1 - \frac{C}{C_{crit}} \right)^m$	[38]	
Wang	$r = \frac{u_{max}}{1 + \left(\frac{C}{K_C}\right)^m}$	[39]	
Liu	$r = \frac{u_{max}K_c}{K_c + C}$	[40]	
Modified Andrews	$r = \frac{\frac{K_C + C}{u_{max}C}}{\frac{K_S + C + \left(\frac{C^2}{K_i}\right)}}$	[41]	
Shukor	$r = v_{max} \left( 1 - \left(\frac{C}{S_m}\right)^n \right)$	(Shukor 2019)	

#### Shukor

#### Fitting of the data

The nonlinear equations were fitted with a Marquardt algorithm using CurveExpert Professional software (Version 1.6). The algorithm searches the best method that minimizes the sum of the squares between predicted and measured values. The software calculates the starting values automatically through via the steepest ascent method.

#### Statistical analysis

The quality of fit of the models to the experimental data was evaluated statistically using the Root-Mean-Square Error (RMSE) (Eqn. 2), adjusted coefficient of determination  $(R^2)$ (Eqn. 3), corrected AICc (Akaike Information Criterion) (Eqn. 4), bias factor (BF) (Eqn. 5) and accuracy factor (AF) (Eqn. 6) as carried out in previous works [42-46].

$$RMSE = \sqrt{\frac{\sum_{i=1}^{n} (Pd_i - Ob_i)^2}{n - p}}$$
 (Eqn. 2)

Adjusted 
$$(R^2) = 1 - \frac{(1 - R^2)(n - 1)}{(n - p - 1)}$$
 (Eqn. 3)

$$AICc=2p+n\ln\left(\frac{RSS}{n}\right)+2(p+1)+\frac{2(p+1)(p+2)}{n-p-2}$$
 (Eqn. 4)

n and p represent the number of data points and the number of parameters, respectively. A model is more likely to be correct if it has the smallest AICc value compared to other models [47].

Bias factor = 
$$10^{\left(\sum_{i=1}^{n} \log \frac{(Pd_i / Ob_i)}{n}\right)}$$
 (Eqn. 5)

Accuracy factor =10

(Eqn. 6)

#### **RESULTS AND DISCUSSION**

Zinc shows an inhibitory effect to the growth of this bacterium and increasing the concentration of this toxicant further inhibited growth with a cessation of growth was observed at the zinc concentration of 6 mgL-1 (Fig. 1). The growth of this bacterium was successfully modelled using the modified Gompertz model with visually acceptable fit to the datapoint was obtained (Fig. 2). The specific growth rates obtained from this modelling exercise was then plugged into the above heavy metal inhibition kinetics models. Two of the models; Liu (Fig. 5) and Andrews (Fig. 6) show poor modelling with the Andrew models exhibiting some problem with the equations as the fitted data needs to pass through the origin, probably due to the  $K_s$  and  $K_i$  terms in the equation, and this might indicate the unsuitability of the Andrews model originally included in the works of Gopinath et al [48] in modelling the effect of heavy metals to the specific growth rate in general.

The best model was Shukor based on the lowest values for RMSE and AICc, highest adjusted correlation coefficient  $(adR^2)$  and values of AF and BF closest to unity (**Table 2**). The parameters obtained from the Shukor model, which are  $S_m$ ,  $\mu_{max}$  and *n* which represent critical heavy metal ion concentration (mg L<sup>-1</sup>), maximum growth rate (h<sup>-1</sup>) and empirical constant values were 6.0 (95%, confidence interval from 5.87 to 6.14), 0.09 (95%, confidence interval of 0.086 to 0.096) and 4.2 (95%, confidence interval from 3.1 to 5.2), respectively.



**Fig. 1**. The effect of increasing concentrations of zinc to the growth of *Pseudomonas* sp. strain DRY15 on SDS. Data represent mean standard deviations (three replicates).



**Fig. 2**. The effect of increasing concentrations of zinc to the growth of *Pseudomonas* sp. strain DRY15 on SDS as fitted to the modified Gompertz model.



Fig. 3. The effect of increasing concentrations of zinc to the maximum specific growth rate of *Pseudomonas* sp. strain DRY15 on SDS as fitted to the Wang model.



Fig. 4. The effect of increasing concentrations of zinc to the maximum specific growth rate of *Pseudomonas* sp. strain DRY15 on SDS as fitted to the Hans-Levenspiel model.



Fig. 5. The effect of increasing concentrations of zinc to the maximum specific growth rate of *Pseudomonas* sp. strain DRY15 on SDS as fitted to the Liu model.



**Fig. 6.** The effect of increasing concentrations of zinc to the maximum specific growth rate of *Pseudomonas* sp. strain DRY15 on SDS as fitted to the Andrews model.



Fig. 7. The effect of increasing concentrations of zinc to the maximum specific growth rate of *Pseudomonas* sp. strain DRY15 on SDS as fitted to the Shukor model.

**Table 2.** Error function analysis of the effect of increasing concentrations of zinc to the maximum specific growth rate of *Pseudomonas* sp. strain DRY15 on SDS as fitted to various secondary models.

Model	р	RMSE	$adR^2$	AF	BF	AICc				
Wang	3	0.01	0.92	1.06	0.98	-38.56				
Levenspiel	3	0.01	0.94	1.05	0.98	-39.08				
Liu	2	0.02	-0.90	1.14	0.92	-36.19				
Andrews	4	0.06	-4.14	2.03	0.49	32.57				
Shukor	3	0.00	0.99	1.02	0.99	-51.71				
Note:										
RMSE	Root mean Square Error									
р	no of parameters									
adR <sup>2</sup>	Adjusted Coefficient of determination									
BF	Bias factor									
AF	Accuracy factor									
AICc	Adjusted Akaike Information Criterion									

In a similar study, zinc inhibits the biodegradation of Congo red by *Pseudomonas* sp. mutant [48] where there was an observable increase in lag period with increasing Zn (II) concentrations. Similarly, the specific growth rate decreasing gradually with increasing Zn (II) concentrations up to 200 mg L<sup>-1</sup> of Zn (II) and later surged rapidly towards zero. In another work, the inhibition constants (K<sub>i</sub>) for the Andrews model in modelling the effect of zinc on the growth rate of *Bacillus* sp. and *Pseudomonas* sp. on monoaromatics such as ethylbenzene, oxylene and toluene ranged from 20 to 26 mg L<sup>-1</sup> [49] indicating that the growth rate on SDS is severely inhibited by zinc. To combat this issue, soil amendments technique such as the addition phosphate, calcium carbonate, manganese oxide, and magnesium hydroxide to reduce the solubility of zinc and to allow bioremediation to proceed [50,51]. Alternatively, immobilization of the SDS-degrading bacterium could offer some protection to combat the toxicity metal ions in general [52].

## CONCLUSION

The biodegradation of SDS by an Antarctic bacterium was found to affect the growth rate on SDS. The best model to study this inhibition to the growth rate was Shukor of which the model predicted accurately the concentration of zinc that caused a complete cessation of growth rate.

#### ACKNOWLEDGEMENTS

This work was supported by the research grants from The Academy of Science Malaysia (ASM) and The Ministry of Science, Technology and Innovation (MOSTI). We thank the Argentinean Institute of Antarctica (IAA) for providing site support during the expedition to Antarctica.

## REFERENCES

- Junfeng Y, Haowen C, Baoling W, Yongqi L. The anion detergent pollution of Antarctic Maxwell Bay and its adjacent sea areas. China Environ Sci. 1998;18(2):151–3.
- George AL. Seasonal factors affecting surfactant biodegradation in Antarctic coastal waters: Comparison of a polluted and pristine site. Mar Environ Res. 2002;53(4):403–15.
- Liwarska-Bizukojc E, Miksch K, Malachowska-Jutsz A, Kalka J. Acute toxicity and genotoxicity of five selected anionic and nonionic surfactants. Chemosphere. 2005;58(9):1249–53.
- Chukwu LO, Odunzeh CC. Relative toxicity of spent lubricant oil and detergent against benthic macro-invertebrates of a west African estuarine lagoon. J Environ Biol. 2006;27(3):479–84.
- Kumar M, Trivedi SP, Misra A, Sharma S. Histopathological changes in testis of the freshwater fish, *Heteropneustes fossilis* (Bloch) exposed to linear alkyl benzene sulphonate (LAS). J Environ Biol. 2007;28(3):679–84.
- Pettersson A, Adamsson M, Dave G. Toxicity and detoxification of Swedish detergents and softener products. Chemosphere. 2000;41(10):1611–20.
- Rosety M, Ribelles A, Rosety-Rodriguez M, Carrasco C, Ordonez FJ, Rosety JM, et al. Morpho-histochemical study of the biological effects of sodium dodecyl sulphate on the digestive gland of the Portuguese oyster. Histol Histopathol. 2000;15(4):1137–43.
- Cserháti T, Forgács E, Oros G. Biological activity and environmental impact of anionic surfactants. Environ Int. 2002;28(5):337–48.
- Pant D, Singh A, Satyawali Y, Gupta RK. Effect of carbon and nitrogen source amendment on synthetic dyes decolourizing efficiency of white-rot fungus, *Phanerochaete chrysosporium*. J Environ Biol. 2008;29(1):79–84.
- Syed M, Mahamood M, Shukor M, Shamaan NA, others. Isolation and characterization of SDS-degrading *Pseudomonas aeruginosa* sp. strain D1. Aust J Basic Appl Sci. 2010;4(10):5000–5011.
- Halmi MIE, Hussin WSW, Aqlima A, Syed MA, Ruberto L, MacCormack WP, et al. Characterization of a sodium dodecyl sulphate-degrading *Pseudomonas* sp. strain DRY15 from Antarctic soil. J Environ Biol. 2013;34(6):1077–82.
- Halmi MIE, Zuhainis SW, Yusof MT, Shaharuddin NA, Helmi W, Shukor Y, et al. Hexavalent molybdenum reduction to Mo-blue by a sodium-dodecyl-sulfate- degrading *Klebsiella oxytoca* strain DRY14. BioMed Res Int. 2013;2013:Article number 384541.

- Shukor MS, Sulaiman MR. Assessment of acetylcholinesterase (AChE) from silver catfish (*Pangasius* sp.) as an assay for organophosphates and carbamates. Biosci Biotechnol Res Asia. 2013;10(1):213–8.
- Masdor N, Abd Shukor MS, Khan A, Bin Halmi MIE, Abdullah SRS, Shamaan NA, et al. Isolation and characterization of a molybdenum-reducing and SDS- degrading *Klebsiella oxytoca* strain Aft-7 and its bioremediation application in the environment. Biodiversitas. 2015;16(2):238–46.
- 15. Maarof MZ, Shukor MY, Mohamad O, Karamba KI, Halmi MIE, Rahman MFA, et al. Isolation and Characterization of a Molybdenum-reducing *Bacillus amyloliquefaciens* strain KIK-12 in Soils from Nigeria with the Ability to grow on SDS. J Environ Microbiol Toxicol. 2018 Jul 31;6(1):13–20.
- Payne WJ, Feisal VE. Bacterial utilization of dodecyl sulfate and dodecyl benzene sulfonate. Appl Microbiol. 1963;11:339–44.
- Roig MG, Pedraz MA, Sanchez JM, Huska J, Tóth D. Sorption isotherms and kinetics in the primary biodegradation of anionic surfactants by immobilized bacteria: II. Comamonas terrigena N3H. J Mol Catal - B Enzym. 1998;4(5–6):271–81.
- Dhouib A, Hamad N, Hassaïri I, Sayadi S. Degradation of anionic surfactants by *Citrobacter braakii*. Process Biochem. 2003;38(8):1245–50.
- Shukor MY, Husin WSW, Rahman MFA, Shamaan NA, Syed MA. Isolation and characterization of an SDS-degrading *Klebsiella* oxytoca. J Environ Biol. 2009;30(1):129–34.
- Chaturvedi V, Kumar A. Isolation of a strain of *Pseudomonas* putida capable of metabolizing anionic detergent sodium dodecyl sulfate (SDS). Iran J Microbiol. 2011;3(1):47–53.
- Chaturvedi V, Kumar A. Diversity of culturable sodium dodecyl sulfate (SDS) degrading bacteria isolated from detergent contaminated ponds situated in Varanasi city, India. Int Biodeterior Biodegrad. 2011;65(7):961–71.
- Chaturvedi V, Kumar A. Presence of SDS-degrading enzyme, alkyl sulfatase (SdsA1) is specific to different strains of Pseudomonas aeruginosa. Process Biochem. 2013;48(4):688–93.
- Shahbazi R, Kasra-Kermanshahi R, Gharavi S, Moosavi-Nejad Z, Borzooee F. Screening of SDS-degrading bacteria from car wash wastewater and study of the alkylsulfatase enzyme activity. Iran J Microbiol. 2013;5(2):153–8.
- Yilmaz F, Icgen B. Characterization of SDS-degrading *Delftia* acidovorans and in situ monitoring of its temporal succession in SDS-contaminated surface waters. Environ Sci Pollut Res. 2014;21(12):7413–24.
- Furmanczyk EM, Kaminski MA, Spolnik G, Sojka M, Danikiewicz W, Dziembowski A, et al. Isolation and characterization of *Pseudomonas* spp. strains that efficiently decompose sodium dodecyl sulfate. Front Microbiol [Internet]. 2017 [cited 2019 Jun 9];8. Available from: https://www.frontiersin.org/articles/10.3389/fmicb.2017.01872/ful 1
- Furmanczyk EM, Kaminski MA, Lipinski L, Dziembowski A, Sobczak A. *Pseudomonas laurylsulfatovorans* sp. nov., sodium dodecyl sulfate degrading bacteria, isolated from the peaty soil of a wastewater treatment plant. Syst Appl Microbiol. 2018 Jul;41(4):348–54.
- Osadebe AU, Onyiliogwu CA, Suleiman BM, Okpokwasili GC. Microbial degradation of anionic surfactants from laundry detergents commonly discharged into a riverine ecosystem. J Appl Life Sci Int. 2018 Apr 3;1–11.
- Gomaa A. Biodegradation of anionic surfactants (sds) by bacteria isolated from waste water in Taif governate. Annu Res Rev Biol. 2018 May 4;26(4):1–13.
- Faria CV de, Delforno TP, Okada DY, Varesche MBA. Evaluation of anionic surfactant removal by anaerobic degradation of commercial laundry wastewater and domestic sewage. Environ Technol. 2019 Apr 3;40(8):988–96.
- Margesin R, Schinner F. Biodegradation of the anionic surfactant sodium dodecyl sulfate at low temperatures. Int Biodeterior Biodegrad. 1998;41(2):139–43.
- Li M, Niu H, Zhao G, Tian L, Huang X, Zhang J, et al. Analysis of mathematical models of *Pseudomonas spp.* growth in palletpackage pork stored at different temperatures. Meat Sci. 2013 Apr 1;93(4):855–64.

- Espeche MC, Tomás MSJ, Wiese B, Bru E, Nader-Macías MEF. Physicochemical factors differentially affect the biomass and bacteriocin production by bovine Enterococcus mundtii CRL1656. J Dairy Sci. 2014;97(2):789–97.
- Halmi MIE, Shukor MS, Johari WLW, Shukor MY. Evaluation of several mathematical models for fitting the growth of the algae *Dunaliella tertiolecta*. Asian J Plant Biol. 2014;2(1):1–6.
- Halmi MIE, Shukor MS, Masdor NA, Shamaan NA, Shukor MY. Evaluation of several mathematical models for fitting the growth of sludge microbes on PEG 600. J Environ Microbiol Toxicol. 2015;3(1):1–5.
- Shen J, Zhang X, Chen D, Liu X, Zhang L, Sun X, et al. Kinetics study of pyridine biodegradation by a novel bacterial strain, *Rhizobium* sp. NJUST18. Bioprocess Biosyst Eng. 2014;37(6):1185–92.
- Çelekli A, Bozkurt H, Dönmez G. Predictive modeling of βcarotene accumulation by Dunaliella salina as a function of pH, NaCI, and irradiance. Russ J Plant Physiol. 2014;61(2):215–23.
- Mansur R, Gusmanizar N, Dahalan FA, Masdor NA, Ahmad SA, Shukor MS, et al. Isolation and characterization of a molybdenumreducing and amide-degrading *Burkholderia cepacia* strain neni-11 in soils from west Sumatera, Indonesia. IIOAB. 2016;7(1):28–40.
- Wang J, Wan W. Kinetic models for fermentative hydrogen production: A review. Int J Hydrog Energy. 2009;34(8):3313–23.
- 39. Wang Y, Zhao Q-B, Mu Y, Yu H-Q, Harada H, Li Y-Y. Biohydrogen production with mixed anaerobic cultures in the presence of high-concentration acetate. Int J Hydrog Energy. 2008 Feb 1;33(4):1164–71.
- Liu Y. A simple thermodynamic approach for derivation of a general Monod equation for microbial growth. Biochem Eng J. 2006;31(1):102-5.
- Andrews JF. A mathematical model for the continuous culture of microorganisms utilizing inhibitory substrates. Biotechnol Bioeng. 1968 Nov 1;10(6):707–23.
- Halmi M, Shukor M, Shukor M. Evaluation of several mathematical models for fitting the growth and kinetics of the catechol-degrading *Candida parapsilopsis*: part 2. J Environ Bioremediation Toxicol. 2014;2(2):53–57.
- Halmi MIE, Shukor MS, Johari WLW, Shukor MY. Mathematical modeling of the growth kinetics of *Bacillus* sp. on tannery effluent containing chromate. J Environ Bioremediation Toxicol. 2014;2(1):6–10.
- Shukor M. Mathematical modelling of the growth of *Klebsiella* pneumoniae on 2-methylquinoline. Bioremediation Sci Technol Res. 2015;3(1):16–19.
- 45. Yakasai MH, İbrahim KK, Yasid NA, Halmi MIE, Rahman MFA, Shukor MY. Mathematical modelling of molybdenum reduction to mo-blue by a cyanide-degrading bacterium. Bioremediation Sci Technol Res. 2016 Dec 31;4(2):1–5.
- 46. Sabullah MK, Rahman MF, Ahmad SA, Sulaiman MR, Shukor MS, Shamaan NA, et al. Assessing resistance and bioremediation ability of *Enterobacter* sp. strain saw-1 on molybdenum in various heavy metals and pesticides. J Math Fundam Sci. 2017 Oct 3;49(2):193–210.
- Burnham KP, Anderson DR. Model Selection and Multimodel Inference: A Practical Information-Theoretic Approach. Springer Science & Business Media; 2002. 528 p.
- Gopinath KP, Kathiravan MN, Srinivasan R, Sankaranarayanan S. Evaluation and elimination of inhibitory effects of salts and heavy metal ions on biodegradation of Congo red by *Pseudomonas* sp. mutant. Bioresour Technol. 2011;102(4):3687–3693.
- Amor L, Kennes C, Veiga MC. Kinetics of inhibition in the biodegradation of monoaromatic hydrocarbons in presence of heavy metals. Bioresour Technol. 2001 Jun 1;78(2):181–5.
- Hettiarachchi GM, Pierzynski GM, Ransom MD. In situ stabilization of soil lead using phosphorus and manganese oxide. Environ Sci Technol. 2000;34(21):4614–9.
- Deeb BE, Altalhi AD. Degradative plasmid and heavy metal resistance plasmid naturally coexist in phenol and cyanide assimilating bacteria. Am J Biochem Biotechnol. 2009;5(2):84–93.
- Halmi MIE, Ahmad SA, Yusof MT, Shukor MY, Syed MA. Entrapment of Mo-reducing bacterium increase its resistance towards heavy metals. Bull Environ Sci Manag. 2013;1(1):11–3.